

Origin of the 28-*pro-R* and 28-*pro-S* Hydrogens in the Biosynthesis of Poriferasterol in *Ochromonas malhamensis*

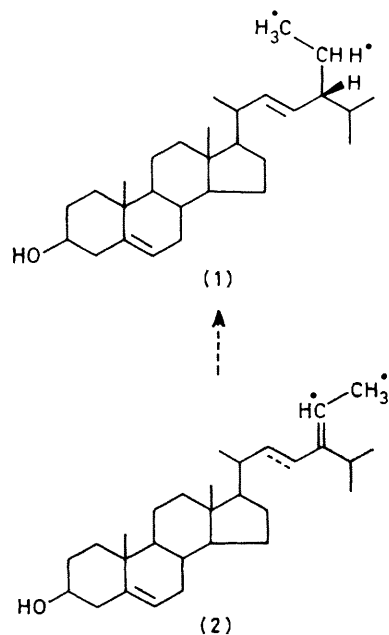
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Summary It is shown, by ^2H n.m.r. spectroscopy, that in the biosynthesis of poriferasterol in *Ochromonas malhamensis* the hydrogen atom at C-28 arising from *S*-adenosylmethionine assumes the 28-*pro-S* position, whereas the hydrogen atom coming from the reducing system assumes the 28-*pro-R* position.

Most C_{29} phyto-sterols are characterized by an ethyl-group at C-24, which arises by a double transmethylation from *S*-adenosylmethionine;¹⁻³ even though the mechanism of this process has been extensively studied,⁴⁻⁹ its stereochemical aspects have been less-well investigated. It is difficult to determine the stereochemistry of the hydrogen atoms at C-28 by the usual tracer techniques (*i.e.* tritium) owing to the lack of chemical and/or biological reactions allowing stereospecific analysis of the side chain. A more promising approach to the solution of these problems seems to be the ^2H n.m.r. technique, provided that there is a reasonable ^2H enrichment in the biosynthetic sample and that the diastereotopic hydrogens at C-28 have different chemical shifts.

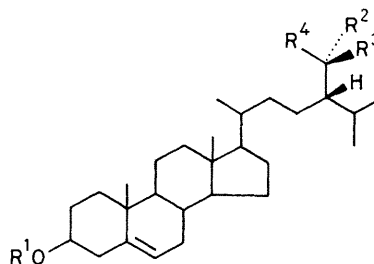
We now report the application of this technique to the study of the stereochemistry of the hydrogen atoms at C-28 in the biosynthesis of poriferasterol (**1**), the major¹⁰ sterol of the chrysophyte alga *Ochromonas malhamensis*.

The formation of this sterol in *Ochromonas malhamensis* occurs *via* an intermediate with a 24(28)-double bond (**2**),⁴ the ethylidene-group of which, arising from *S*-adenosylmethionine, is subsequently reduced to the 24-ethyl-group (Scheme 1). This mechanism implies that poriferasterol



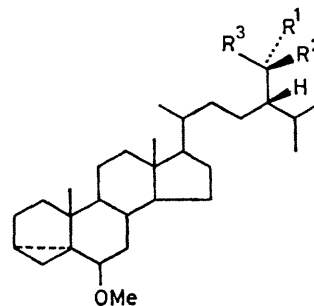
SCHEME. H* = Hydrogen from methionine.

ferasterol which is biosynthesized in *Ochromonas malhamensis* from L-[methyl- $^2\text{H}_3$]methionine will contain a ^2H atom at C-28 arising from methionine and a ^1H atom introduced by the $\Delta^{24(28)}$ -reductase present in the alga. To determine the stereochemistry of the above atoms by ^2H n.m.r. spectroscopy, we had to know the chemical shift values of the diastereotopic 28- H_R and 28- H_S hydrogens. These were measured on model compounds (24*S*,28*S*)-[28- ^2H]- and (24*S*,28*R*)-[28- ^2H]- β -acetoxystigmast-5-ene (**3**) and (**4**), which were synthesized as follows: (24*S*)-6 β -methoxy-3 α ,5-cyclostigmastan-28-one (**5**)¹¹ was reduced with LiAl^2H_4 and the diastereoisomeric deuteriated alcohols (**6**) and (**7**) obtained were separated by repeated preparative t.l.c. and their configurations at C-28 assigned¹¹ by Horeau's procedure. The alcohols (**6**) and (**7**) were then transformed into (**3**) and (**4**) by LiEt_3BH reduction of the mesylates¹¹ and by aceto!ysis.¹²



- (3) $\text{R}^1 = \text{Ac}$; $\text{R}^2 = ^2\text{H}$; $\text{R}^3 = \text{H}$; $\text{R}^4 = \text{Me}$
 (4) $\text{R}^1 = \text{Ac}$; $\text{R}^2 = \text{H}$; $\text{R}^3 = ^2\text{H}$; $\text{R}^4 = \text{Me}$
 (8) $\text{R}^1 = \text{Ac}$; $\text{R}^2 = ^2\text{H}$; $\text{R}^3 = \text{H}$; $\text{R}^4 = \text{C}^2\text{H}_5$; Δ^{22}
 (9) $\text{R}^1 = \text{R}^3 = \text{H}$; $\text{R}^2 = ^2\text{H}$; $\text{R}^4 = \text{C}^2\text{H}_5$; Δ^{22}
 (11) $\text{R}^1 = \text{Ac}$; $\text{R}^2 = ^2\text{H}$; $\text{R}^3 = \text{H}$; $\text{R}^4 = \text{C}^2\text{H}_5$

^2H N.m.r. analysis of the above model compounds (see Table) showed a difference between the chemical shifts of $^2\text{H}_R$ and $^2\text{H}_S$ sufficiently large to determine the position assumed by the 28- ^2H atom in the biosynthetic poriferasterol. To obtain this compound, *Ochromonas mal-*



- (5) $\text{R}^1, \text{R}^2 = \text{O}$; $\text{R}^3 = \text{Me}$
 (6) $\text{R}^1 = \text{OH}$; $\text{R}^2 = ^2\text{H}$; $\text{R}^3 = \text{Me}$
 (7) $\text{R}^1 = ^2\text{H}$; $\text{R}^2 = \text{OH}$; $\text{R}^3 = \text{Me}$
 (10) $\text{R}^1 = ^2\text{H}$; $\text{R}^2 = \text{H}$; $\text{R}^3 = \text{C}^2\text{H}_5$; Δ^{22}

hamensis, 933/1A, Cambridge Culture Collection, was cultured at 27 °C in 20 ml batches¹³ in the presence of L-[methyl-²H₃]methionine (99% ²H)⁴ with continuous shaking. After two days the cells from ten flasks were centrifuged and saponified with 10% KOH in 80% aq. EtOH under reflux for 1 h. The unsaponifiable material (560 mg), extracted with diethyl ether, was chromatographed on silica gel to yield 93 mg of crude sterol fraction; acetylation and purification on 25% AgNO₃/SiO₂ yielded 65 mg of deuteriated poriferasteryl acetate (**8**) (checked by g.l.c., SE-30 2.5%, T_c 225 °C and OV-17 1%, T_c 260 °C, 98% pure). Hydrolysis of the compound (**8**) with methanolic KOH and crystallization from methanol afforded 57 mg of deuteriated poriferasterol (**9**) which showed, by mass spectrometry, the presence of the expected four deuterium atoms in the side chain (34.7% ²H₀, 22.9% ²H₁, 24.9% ²H₂, 17.5% ²H₃, 40.4% ²H enrichment at C-28).

The tetradeuteriated (**9**) was transformed, using the method of Steele and Mosettig,¹⁴ into (24*R*)-[²H₄]6β-methoxy-3α,5-cyclostigmast-22-ene (**10**). Hydrogenation with 10% Pd on carbon and rearrangement with zinc acetate in boiling acetic acid¹² afforded 32 mg of tetradeuteriated (24*S*)-3β-acetoxystigmast-5-ene (**11**), the ¹H n.m.r. spectrum of which was indistinguishable from that of an authentic sample. The ²H n.m.r. spectrum of the tetradeuteriated compound (**11**) (see Table) showed two peaks, the area ratio of which was 3:1; the former peak, at 0.81 p.p.m., corresponded to the trideuteriated C-29 methyl-group and the latter, corresponding to a single deuterium

atom, was found at 1.13 p.p.m., a value essentially identical with that of (24*S*,28*S*)-[28-²H]-3β-acetoxystigmast-5-ene (**3**).

TABLE.

Compound	Chemical shifts values in p.p.m. ^a	
	28- ² H	29-C ² H ₃
(3)	1.19	—
(4)	1.33	—
(11)	1.13	0.81

^a 'Resolution-enhanced' ²H-n.m.r. spectra recorded on a Varian XL-200 instrument at 30.7 MHz in CHCl₃ solution using CDCl₃ at 7.24 p.p.m. as the internal standard.

These results indicate that in the biosynthesis of poriferasterol in *Ochromonas malhamensis* the hydrogen at C-28 arising from S-adenosylmethionine assumes the 28-*pro-S* position, whereas the hydrogen coming from the reducing system assumes the 28-*pro-R* position. Moreover, the absence of any detectable signal due to the 28-*R* deuterium, coupled with the known stereospecificity of the biological reduction processes, strongly suggests that only one of the *E*- and *Z*-24(28)-ethylidene sterols, which is transformed into poriferasterol in *Ochromonas malhamensis*,¹⁵ is the real biosynthetic intermediate.

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¹ E. Lederer, *Quart. Rev., Chem. Soc.*, 1969, **23**, 453.

² L. J. Goad and T. W. Goodwin, *Prog. Phytochem.*, 1972, **3**, 113.

³ L. J. Goad, F. F. Knapp, J. R. Lenton, and T. W. Goodwin, *Lipids*, 1974, **9**, 582.

⁴ A. R. H. Smith, L. J. Goad, T. W. Goodwin, and E. Lederer, *Biochem. J.*, 1967, **104**, 56c.

⁵ J. R. Lenton, L. J. Goad, and T. W. Goodwin, *Phytochemistry*, 1975, **14**, 1523.

⁶ Y. Tomita, A. Uomori, and H. Minato, *Phytochemistry*, 1970, **9**, 555.

⁷ Y. Tomita, A. Uomori, and E. Sakurai, *Phytochemistry*, 1971, **10**, 573.

⁸ R. Ellouz and M. Lenfant, *Eur. J. Biochem.*, 1971, **23**, 544.

⁹ K. H. Raab, N. J. de Souza, and W. R. Nes, *Biochim. Biophys. Acta*, 1968, **152**, 742.

¹⁰ M. C. Gershengorn, A. R. H. Smith, G. Goulston, L. J. Goad, T. W. Goodwin, and T. H. Haines, *Biochemistry*, 1968, **7**, 1698.

¹¹ G. Busca, F. Nicotra, F. Ronchetti, and G. Russo, *Gazz. Chim. Ital.*, 1978, **108**, 665.

¹² G. D. Aderson, T. J. Power, C. Djerassi, and J. Clardy, *J. Am. Chem. Soc.*, 1975, **97**, 388.

¹³ S. Aaronson and H. Baker, *J. Protozool.*, 1959, **6**, 282.

¹⁴ J. A. Steele and E. Mosettig, *J. Org. Chem.*, 1963, **28**, 571.

¹⁵ F. F. Knapp, L. J. Goad, and T. W. Goodwin, *Phytochemistry*, 1977, **16**, 1683.